



**What determines the sensitivity?
– the numbers of scans and sampling –**

IPR-seminar

**Practical Aspects of Non-uniform Sampling
in Multi-dimensional NMR Spectroscopy
and Application for Biological Systems**

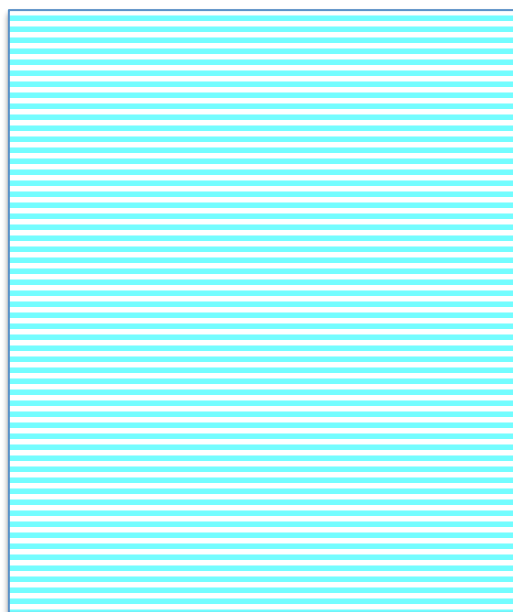
**2014 June 19th (Thr)
Nagoya Univ.**

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**I just want to increase
the number of scans (NS) and
the number of total sampling data (TD₁)
to get 3D spectra of higher sensitivity and
resolution.**

**But, to do this, the experimental time would
extend to 7 days!**

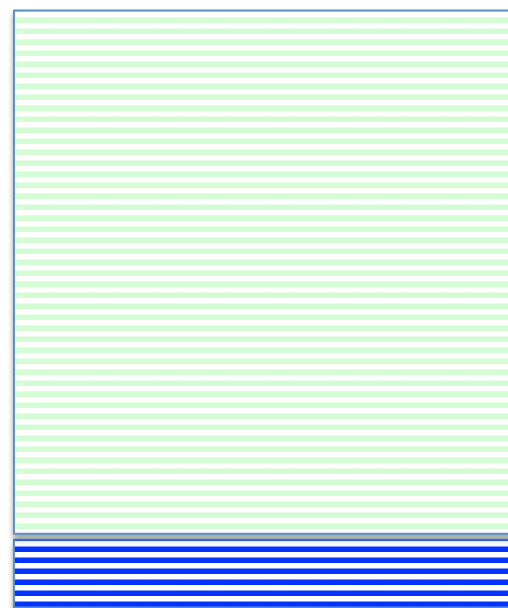
I tried decreasing TD of the ^{15}N indirect dimension, and at the same time, increasing NS, so that the experimental time was conserved.



^{15}N interferogram

^1H FID

NS = 1
TD = 2,560



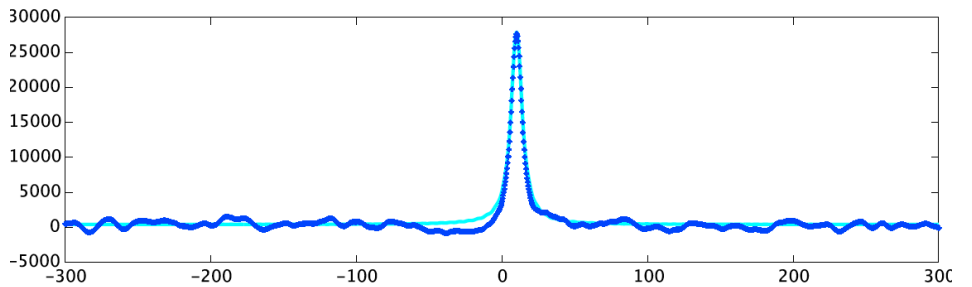
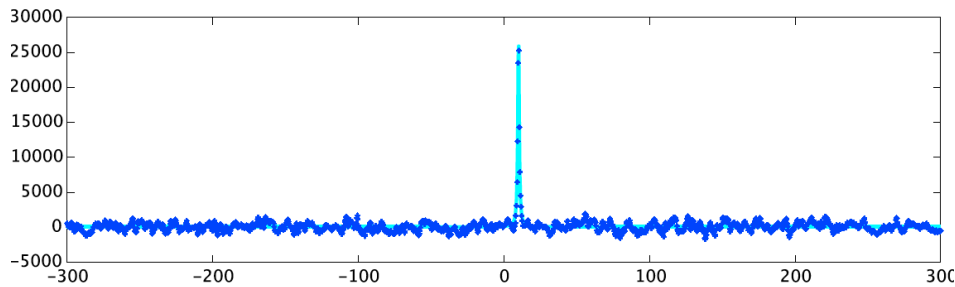
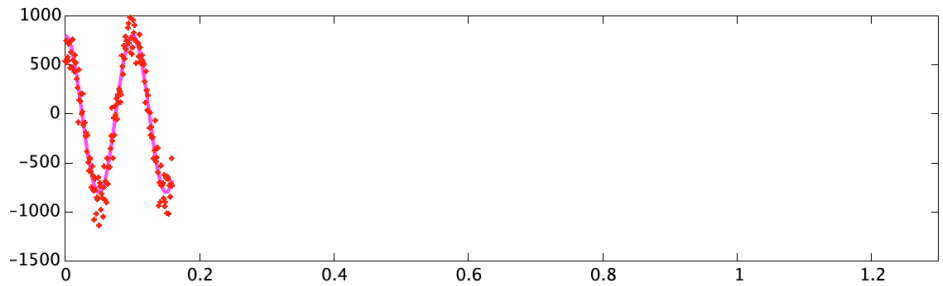
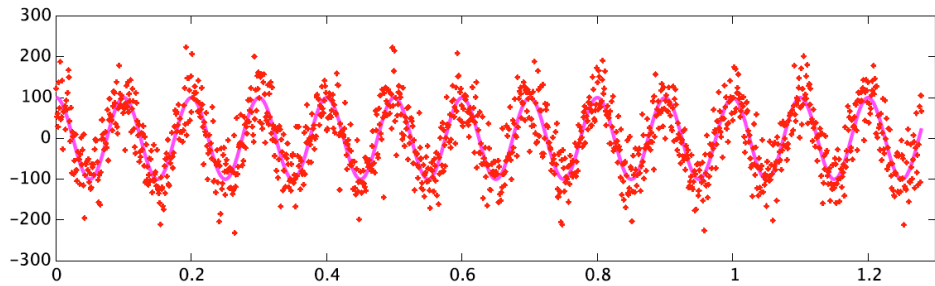
^{15}N interferogram

^1H FID

NS = 8
TD = 320

Let's assume that the interferogram be sampled in the constant-time manner (no decay).

The noise of the interferogram (NS=8) decreased to $1/\sqrt{8} = 35\%$.



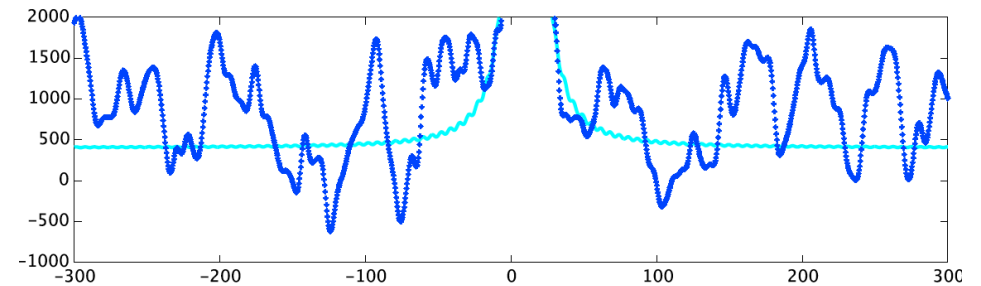
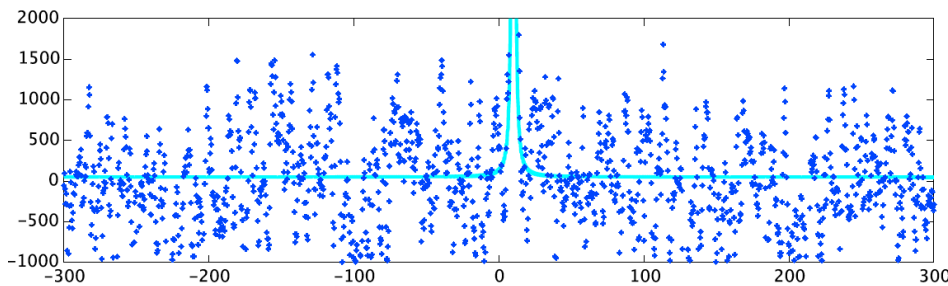
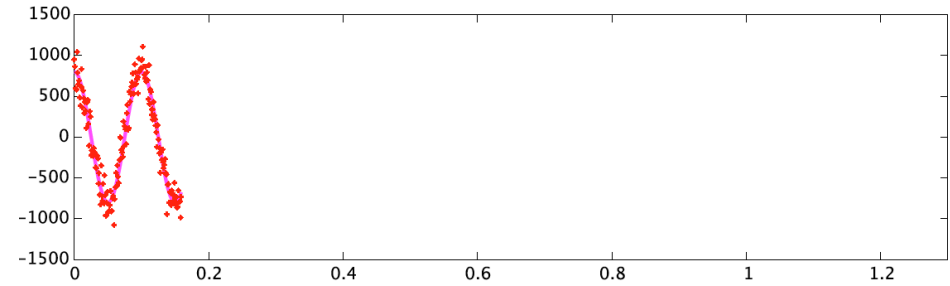
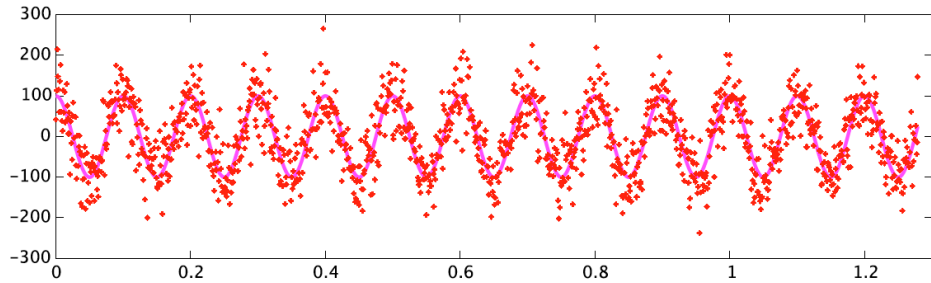
NS = 1
TD = 2,560

sharper!

NS = 8
TD = 320

broadened ...

Let's magnify the spectra to compare them more carefully.



NS = 1
TD = 2,560

spectral noise : 578

NS = 8
TD = 320

spectral noise : 584

The peak and noise both broadened, but S/N was the same!



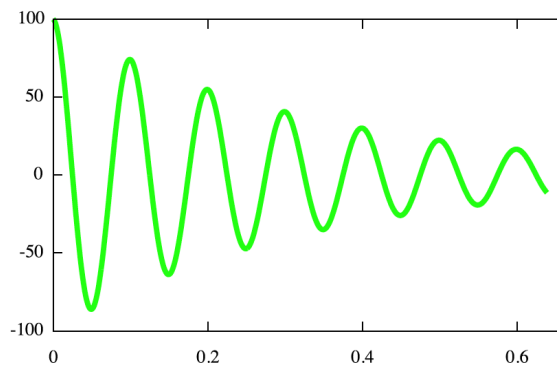
What is sensitivity ?

Signal intensity (S) = the area of FID (interferogram) after modification with a window function.

$$\text{Sensitivity (S/N)} \propto \sqrt{N_s} \cdot \sqrt{T_d} \propto \sqrt{\text{expt}}$$

As far as the *expt* is kept at constant, S/N does not change.

$$N_s = 8$$
$$T_D = 256$$

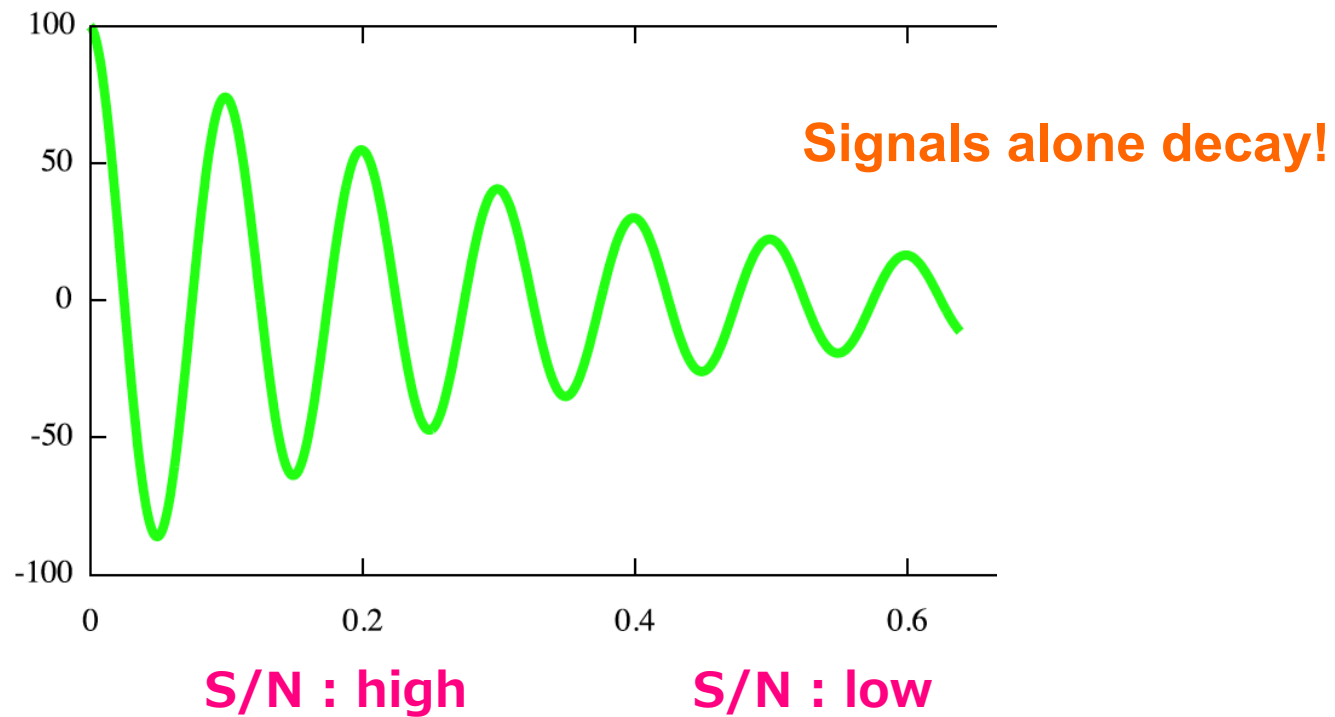


$$N_s = 16$$
$$T_D = 128$$

Practically,

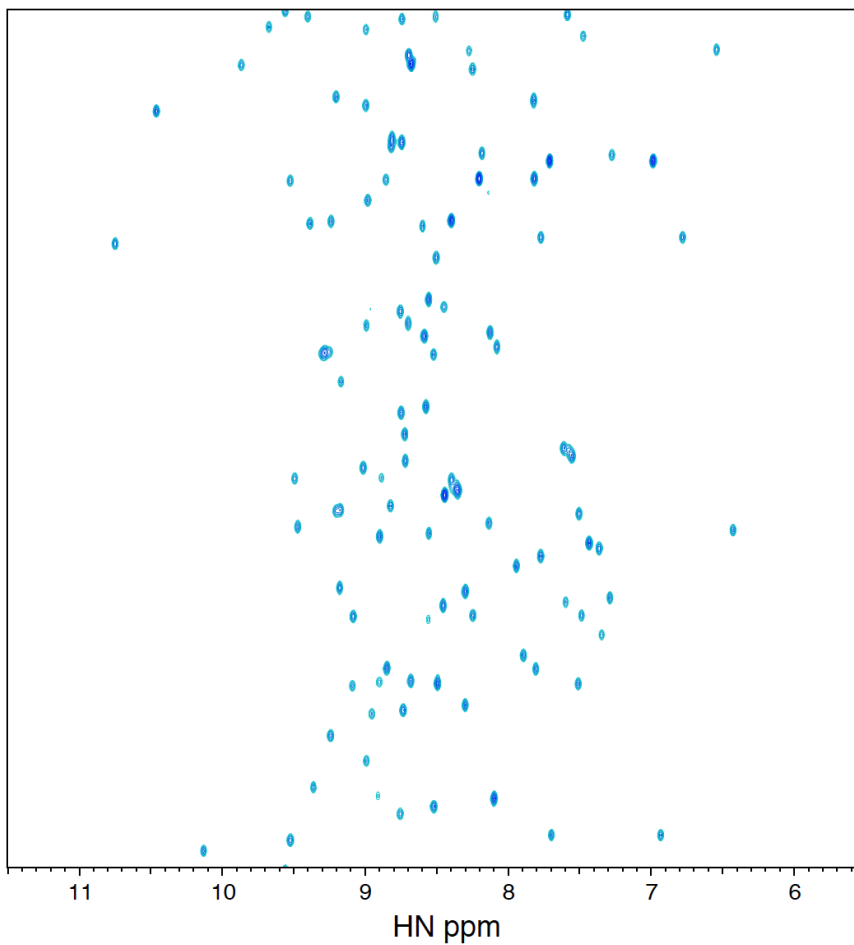
the spectrum (NS=8, sampling=16) : higher S/N

the spectrum (NS=2, sampling=64) : lower S/N



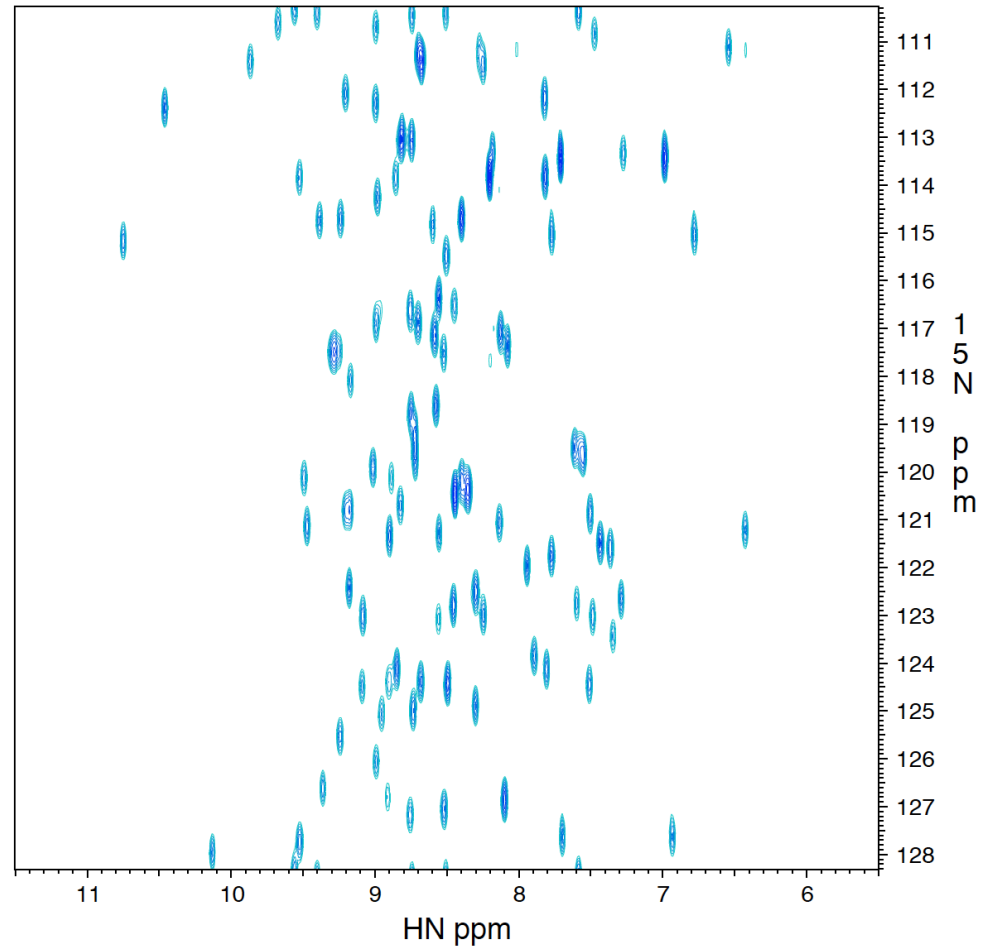
The noise level is the same throughout the interferogram irrespective of relaxation.

Each measurement took 30 min.



NS = 4
TD = 256

S/N → 304

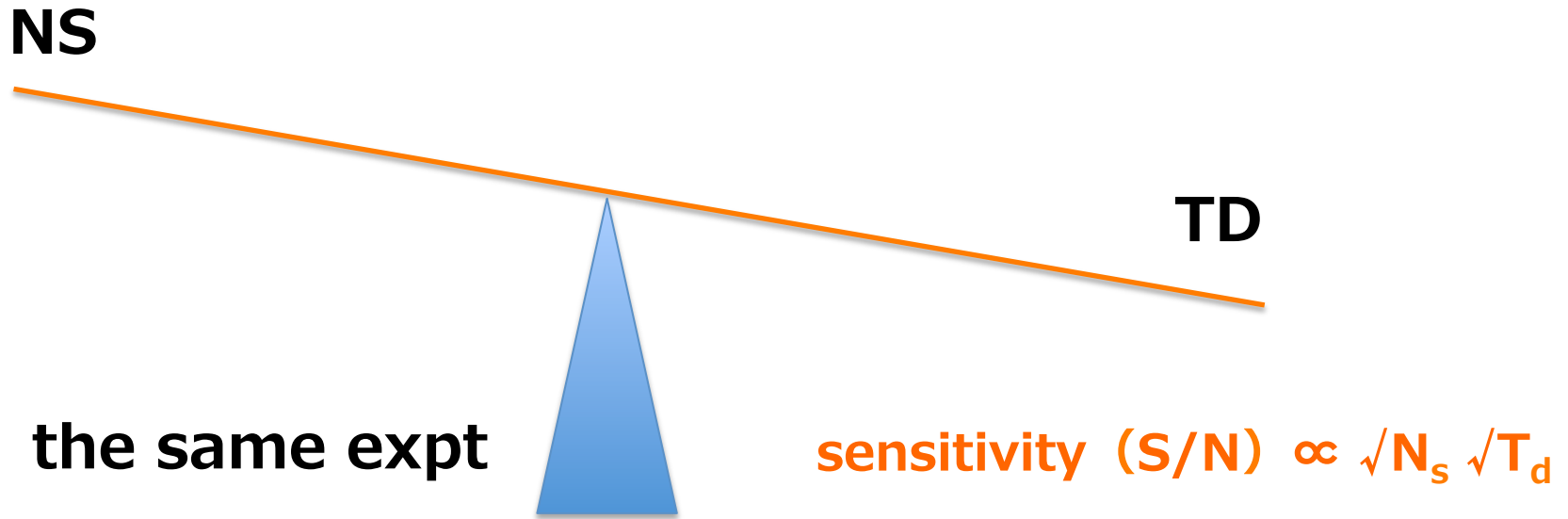


NS = 16
TD = 64

S/N → 335

Slightly higher S/N
owing to ^{15}N T_2 relaxation. →

Any good way to enhance the resolution without extending expt?

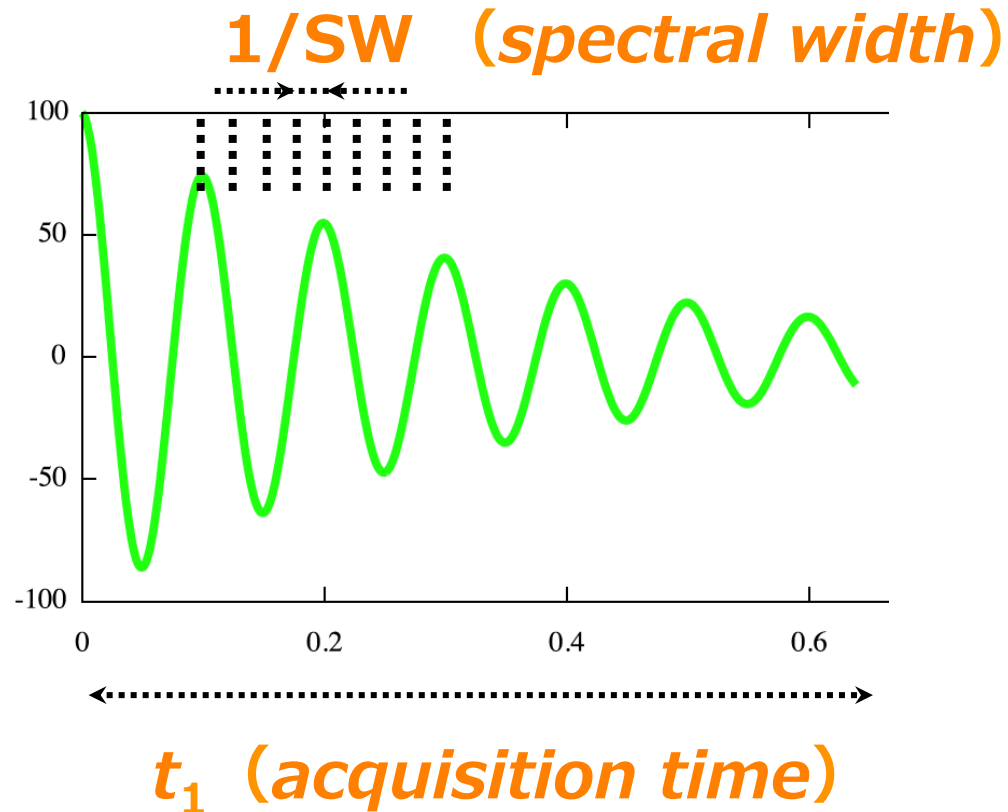


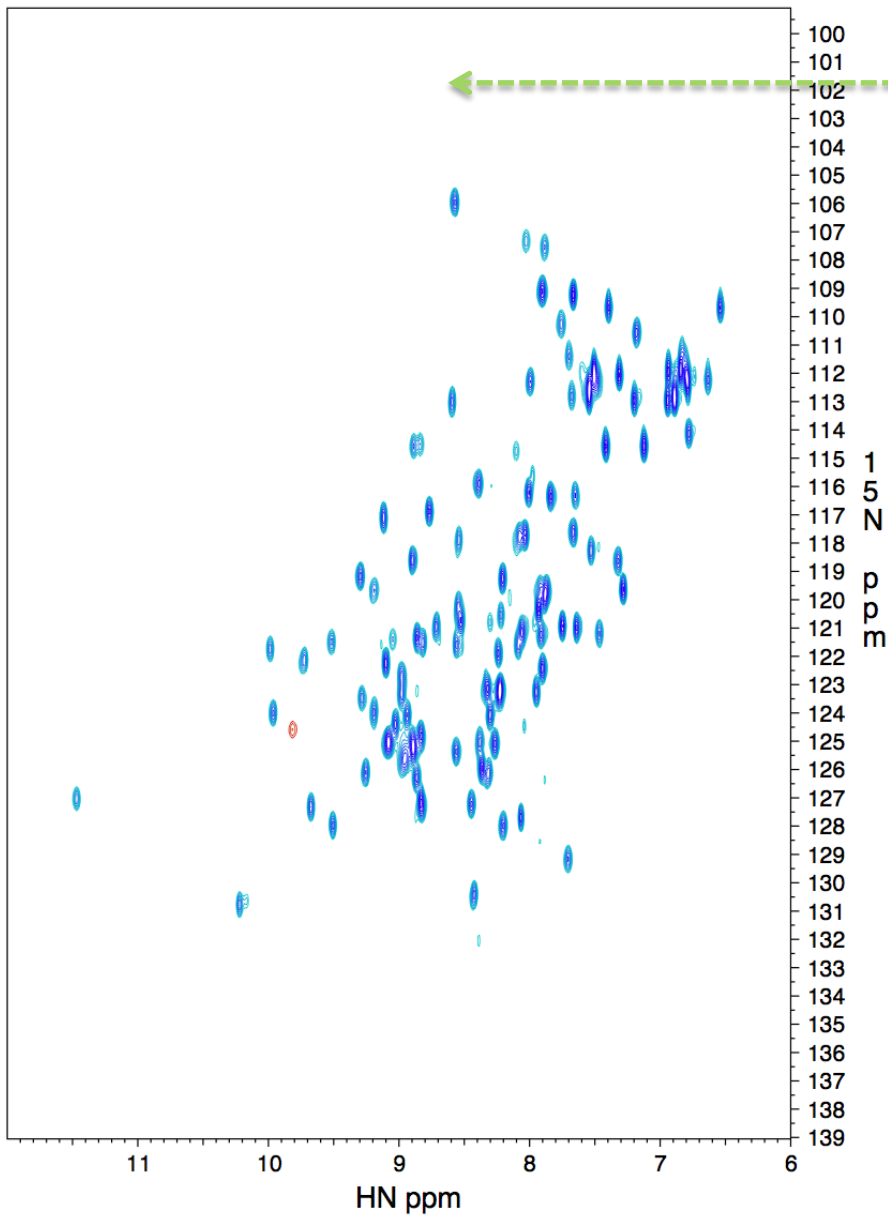
relaxation

- NS: min. and TD: max. → not enough phase cycles
- sampling t_1 non-uniformly = NUS
- just increasing $\Delta t_1 = i.e.,$ narrowing SW

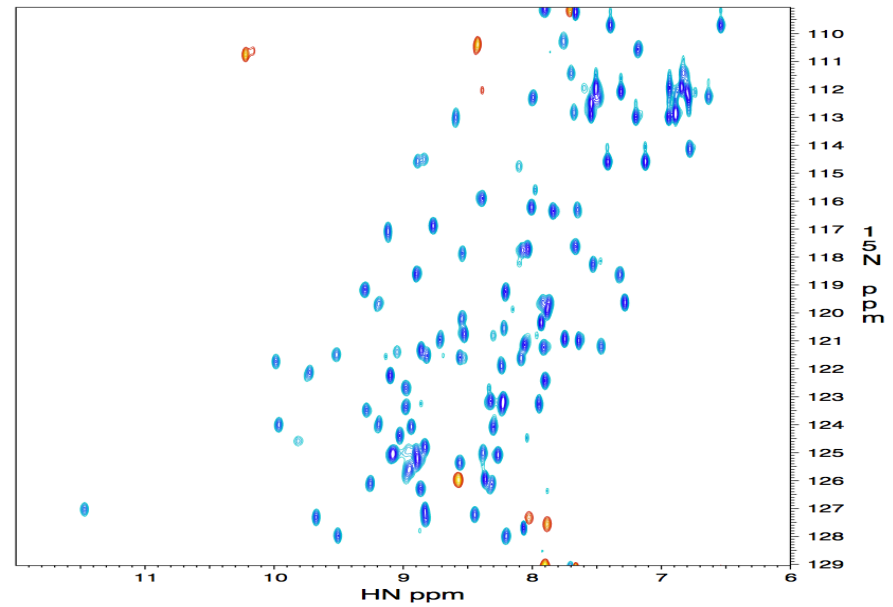
narrower spectral width → higher resolution

$$\text{acquisition time} = \frac{\text{point number}}{\text{spectral width}} = \frac{1}{\text{resolution}}$$

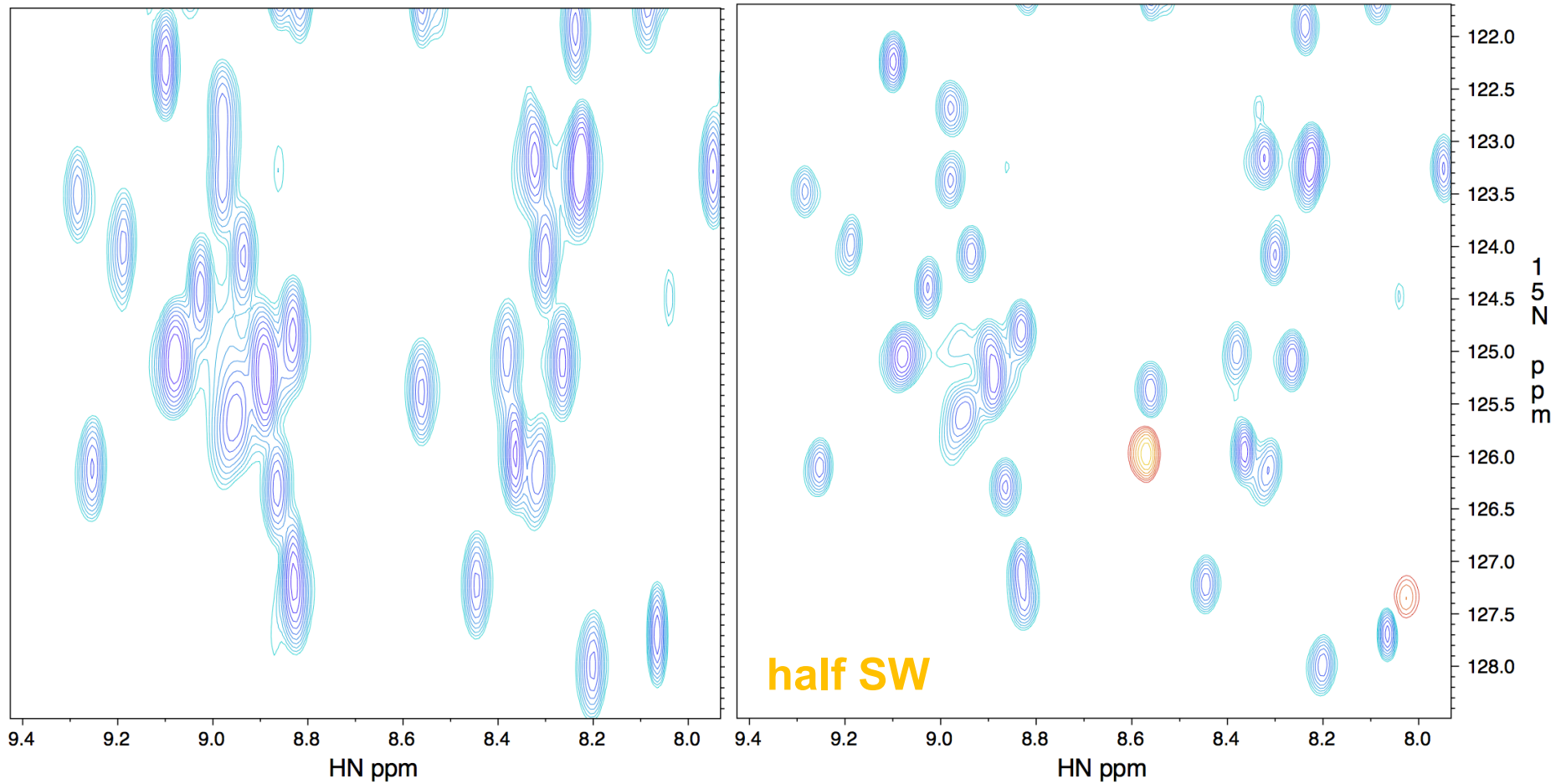




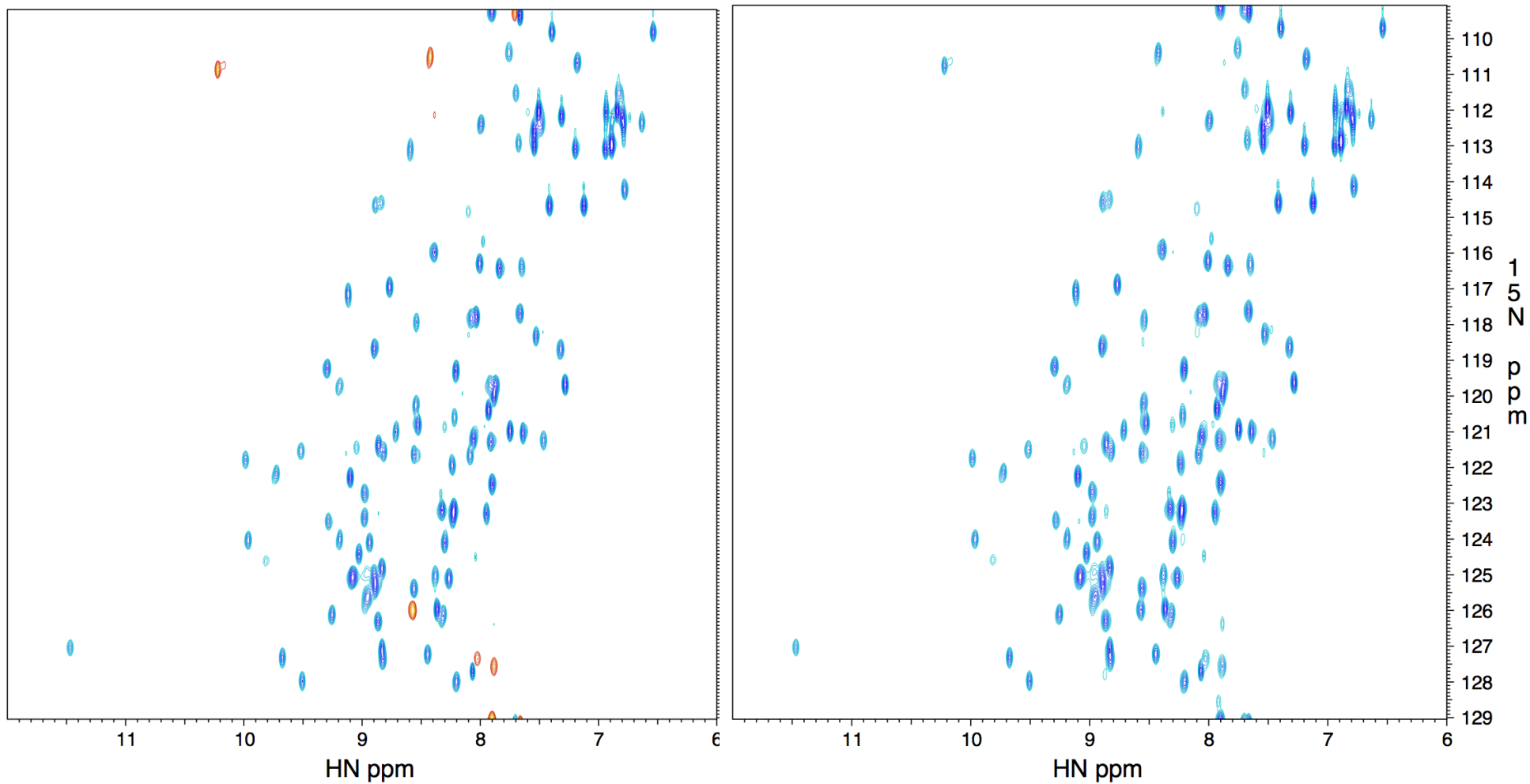
no need to observe these areas (waste of time!)



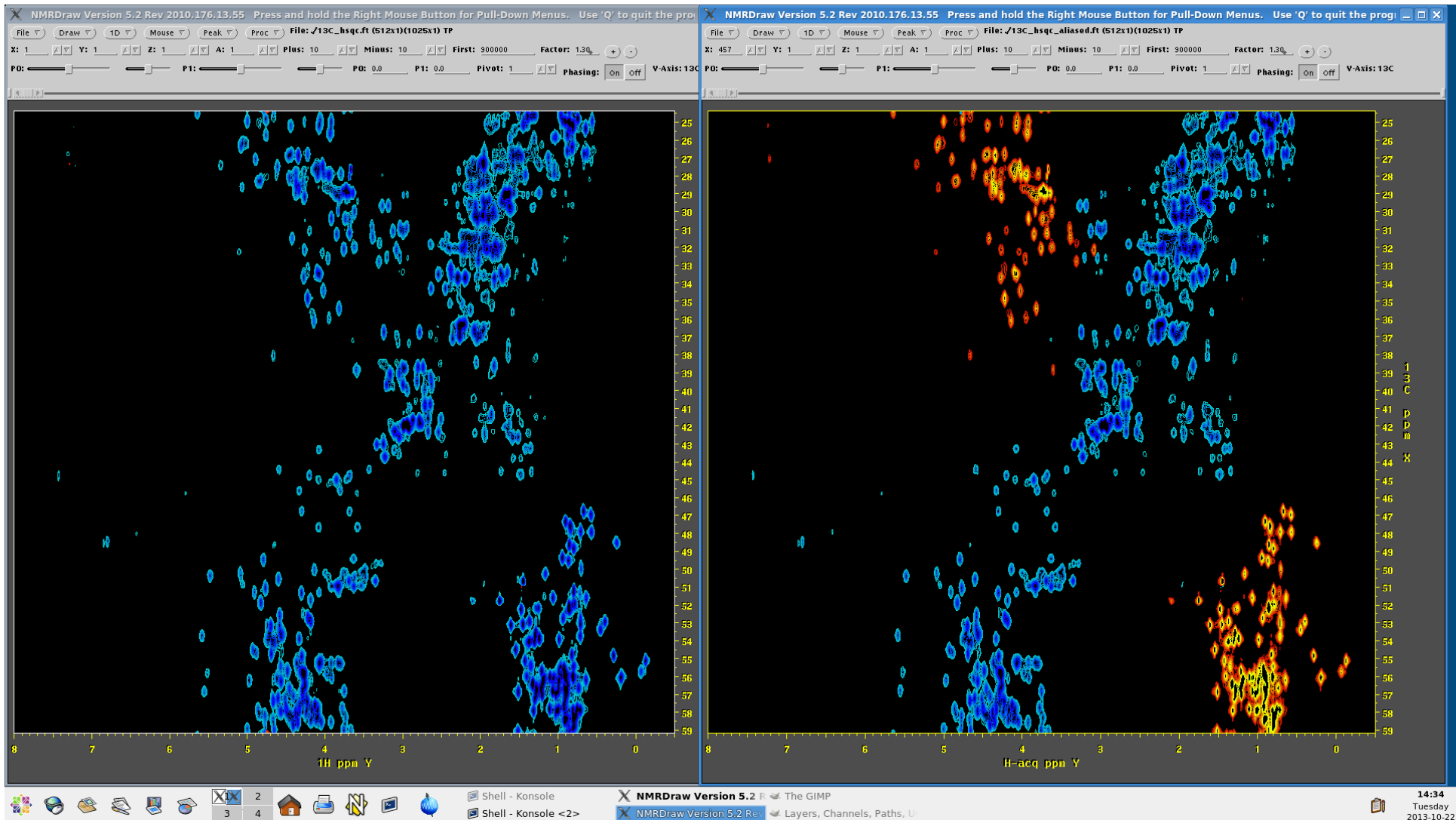
A higher resolution after the same measurement time



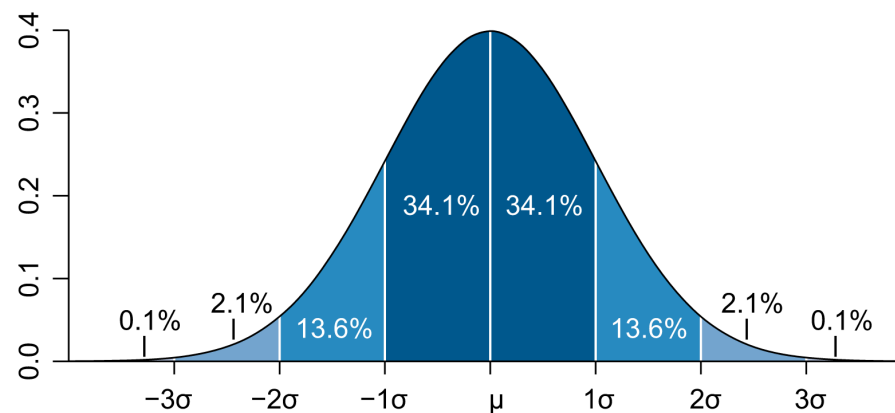
If SW of ω_1 and ω_2 each is reduced by half in 3D, expt can be shortened to $\frac{1}{4}$ (or S/N can be enhanced twice).



if $t_1(0)$ starts with $\Delta t_1/2$, aliased peaks appear negative.
 Let's narrow ¹⁵N SW as much as possible (protein NMR).
 Never narrow ¹H_N, ¹H_C SW (FID)! ... cost of ¥ 6,000,000.-



Important spectra in protein NMR, such as 3D ¹³C-edited NOESY, can be measured with a higher sensitivity by simply utilizing aliasing.



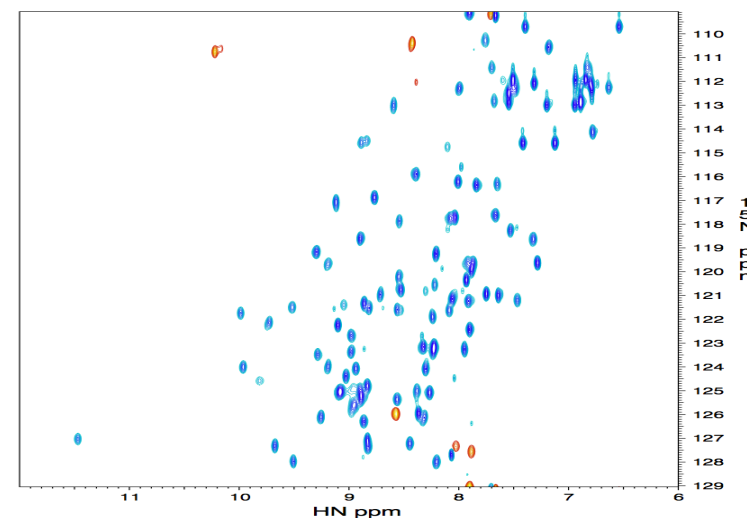
^{13}C SW

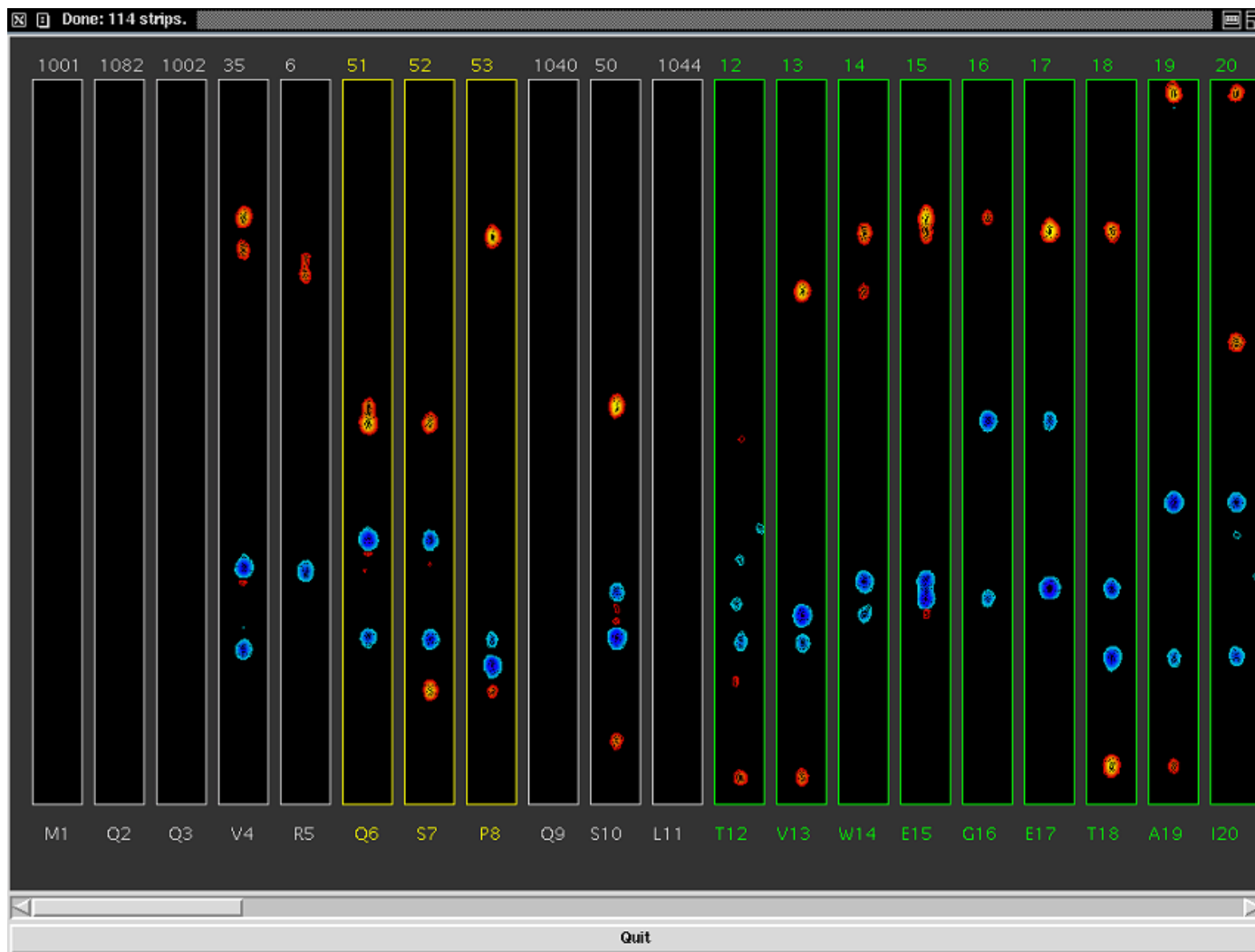
In principle, SW of $\pm 1.5 \sigma$ is enough, since aliased peaks do not appear beyond the center (99.7%).

Just in case, $\pm 2.0 \sigma$ is safer.

$^{13}\text{C}_\alpha$: $176.28 \text{ ppm} \pm 8.06 (3\sigma)$
→ 11 ppm (2σ) is OK.

$^{13}\text{C}_\alpha$: $56.335 \text{ ppm} \pm 15.27 (3\sigma)$
→ 21 ppm (2σ) is OK.





You can assign the protein main-chains sequentially without picking up peaks. You just only need to arrange strips by matching patterns on your *iPad*.

<http://www.nmrscience.com/nmrpipe.html> (NMRWish in F. Delaglio's NMRPipe HP)